

and $f(\vec{p}_i, \vec{h}_j)$ is a formula, describing reflections; d_1 and d_2 are quadratic, or robust, or entropy metrics. At the same time \vec{h}_j^* , computed while minimizing R_j , index j th reflections. V and N are volumes of a phase and total number of reflections at tried parameter vectors.

The minimization of (1) with respect to \vec{p}_i is carried out by trying parameter values from a assumed region, and refining them by analytical fitting, which, in case of convergence, gives the best estimate, or, if initial guesses are good enough, by analytical fitting.

Reference: Zlokazov V.B. Comp.Phys.Comm.,1995,v.85,p.415-422.

PS02.06.21 PHASING THE CHOLERA TOXIN ELECTRON DIFFRACTION DATA USING THE MAXIMUM ENTROPY-LIKELIHOOD METHOD WITH NON-CRYSTALLOGRAPHIC SYMMETRY IMPOSED IN THE MICE COMPUTER PROGRAM. W.N. Nicholson and C.J. Gilmore, Department of Chemistry, University of Glasgow, Glasgow G12 8QQ, Scotland U.K.

We have already reported our experience with applying the maximum entropy-likelihood method to phasing the two-dimensional projection data for cholera toxin (Gilmore & Nicholson (1995). Transactions American Crystallographic Association, 31 In press.). We have been working with a 2-d data set for which 56 unique image phases are available at 8.8Å resolution, and for which a further 1417 diffraction intensities extend to 4Å. The problem has been:

1. To phase the 4Å data from the 56 known phases.
2. To impose 5-fold non-crystallographic symmetry on the projection.
3. To impose envelope and solvent flattening constraints.

The maximum entropy-likelihood program (MICE) has been modified to carry out these steps. Using it we have shown that the likelihood criterion to is an accurate and reliable predictor of:

4. The effective number of atoms in the unit cell.
5. The centroid coordinates for the 5-fold non-crystallographic axes.
3. The envelope radius.

We are now extending the procedure to phase data derived from a series of electron diffraction patterns derived from a limited set of specimen tilts. The extension of two-dimension phase information into three is non-trivial especially when so little a priori phase information is available, and will be discussed in detail.

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Metalloproteins X-Ray & EXAFS Studies

MS02.07.01 EXAFS STUDIES OF NITROGENASE AND RELATED SYSTEMS: AN OVERVIEW. Keith O. Hodgson, Department of Chemistry Stanford University, Stanford, CA 94305

X-ray absorption edge and extended fine structure (EXAFS) studies contributed significantly to the early definition of particularly the Mo-Fe-S containing cofactor (FeMoco) of the MoFe protein of the nitrogenase system both within and outside the protein (in its isolated form). The x-ray crystal structure results have now defined the overall protein structure and the relationships and structures of the metal-containing prosthetic groups in the resting state. However, much remains to be learned about how the electronic structure of the FeMoco mediates electron transfer and dinitrogen reduction and XAS spectroscopy is now being used to probe this question. Edge studies provide information on electronic structure including covalency and electronic distribution as a function of oxidation state. EXAFS results incorporating multiple scattering analyses provide a specific probe of the metrical details (including longer range distances) and in some cases geometry of the FeMoco. This talk will review some of these recent advances as they provide insights into the structure/function of the nitrogenase system.

MS02.07.02 EXAFS AND X-RAY STUDIES OF B₁₂ MODEL COMPOUNDS. Christoph Kratky, Institut für physikalische Chemie, A-8010 Graz - Austria.

The chemical and structural complexity of the B₁₂ coenzymes (5'-desoxyadenosyl cobalamin and methyl cobalamin) and their biologically relevant reactivities (Co-C bond homolysis and Co-C bond heterolysis, respectively) has been a puzzle ever since the elucidation of their structures by Dorothy Hodgkin's group more than three decades ago. For many years, the focus of structural and chemical research lay on the isolated cofactor or cofactor analogue, with special emphasis on the cobalt center and its coordination environment.

Along these lines, we have determined crystal structures of a number of cobalamins with different α - and β -substituents. Using synchrotron radiation in combination with imaging plate detectors for some of these analyses, we were able to collect very accurate and comprehensive data sets, which permitted structure refinement to a level of precision comparable to a well-determined small-molecule crystal structure. From the combined structural data of about 20 cobalamin crystal structures, correlations between several characteristic intra-molecular deformation parameters (upward-folding of the corrin ring, axial Co-N distance, orientation of the dimethylbenzimidazole base) can be established and used to estimate the relative "stiffness" of each of these deformation modes.

In recent years, the B₁₂ field has advanced dramatically as a result of the elucidation of the first crystal structures of proteins binding a B₁₂ cofactor (B₁₂ binding domain of methionine synthase; methylmalonyl CoA mutase). In both proteins, a protein-derived histidine-imidazole occupies the α -axial coordination of the cobalt center, replacing the dimethylbenzimidazole base occupying this position in solutions of the isolated cofactor under physiological conditions. Thus, it appears that some of the above structural correlations refer to a biologically irrelevant cofactor constitution.

For a number of representative compounds, we have also collected X-ray absorption spectra as a basis for the interpretation of EXAFS spectra of cobalamins in "non-crystalline" environments (e.g. cobalamins in solution and bound to a protein). Thus, by a comparison of the spectra of Aquocobalamin perchlorate in solution and in crystalline form, we could show that there is no detect-