

and UAA (ochre). These stop codons are occasionally recoded to an amino acid by specialized transfer RNAs (tRNAs) called suppressor tRNAs. In all organisms, the UGA codon is recoded to selenocysteine (Sec; the 21st amino acid), depending on the downstream enhancer sequence SECIS, through multi-step Sec synthesis on tRNA^{Sec} (opal suppressor). In methanogenic archaea and some anaerobic eubacteria, the UAG codon is recoded to pyrrolysine, the 22nd amino acid, through direct acylation of amber suppressor tRNA^{Pyl} by pyrrolysyl-tRNA synthetase (PylS). The tRNA^{Pyl} suppressor has an unusual structure, lacking most of the consensus features of canonical tRNAs, analogous to mammalian mitochondrial tRNAs. It is thus unclear how such unusual tRNAs function in the ribosome and how PylS discriminates between suppressor and canonical tRNAs. Here we present the crystal structure of the PylS/tRNA^{Pyl} complex from *Desulfotobacterium hafniense* at 3.1 Å resolution. The suppressor tRNA^{Pyl} has an unusual minimal core structure formed by a non-standard D-loop, a TYC-loop and a variable loop, which are reorganized to mimic the canonical L-shape to function normally in the ribosome. PylS recognizes the unusual compact core structure by steric compatibility, which enables discrimination of the suppressor from the other canonical tRNAs.

Keywords: tRNA, aminoacyl-tRNA synthetase, stop codon

MS.08.3

Acta Cryst. (2008). A64, C26

DNA transfer machines

Miquel Coll

Institute for Reserach in Biomedicine & Institut de Biologia Molecular de Barcelona, Structural & Computational Biology, Baldiri Reixac 10-12, Barcelona, Barcelona, 08028, Spain, E-mail : miquel.coll@irbbarcelona.org

Whatever the route used, horizontal gene transfer, a rapid way for bacterial evolution, requires sophisticated protein machinery to enable DNA to cross the cell envelope barriers. Increased antibiotic resistance among pathogens is a troubling consequence of this microbial capacity. Mechanisms leading to horizontal gene transfer in bacteria are categorized into transduction, transformation and conjugation. Transduction occurs via bacteriophages, which may incorporate portions of the host bacterial DNA and introduce them into newly infected hosts. Natural transformation consists of the uptake of naked DNA from the environment. Finally, conjugation is the unidirectional transfer of ssDNA of conjugative plasmids from a donor to a recipient cell. The conjugation system can be divided into two discrete specialized modules: the relaxosome, which triggers plasmid DNA processing and replication, and a type IV secretion system, which impels protein and single-stranded DNA through the membranes. In addition, a coupling protein, which links these two modules, and a number of ancillary proteins are required. Over recent decades, research efforts in the field have clarified many aspects of the system. In particular, structural biology is providing details of the molecular architecture of several of the components involved. Russi, S., Boer, R. & Coll, M. (2008). Molecular Machinery for DNA Translocation in Bacterial Conjugation. In *Plasmids: Current Research and Future Trends*. Horizon Scientific Press, Londres.

Keywords: protein-DNA complexes, DNA translocation, bacterial conjugation

MS.08.4

Acta Cryst. (2008). A64, C26

RNA-protein interactions in the U4 snRNP core domain

Jade Li, Adelaine K W Leung, Kiyoshi Nagai

MRC Laboratory of Molecular Biology, Structural Studies Division, Hills Road, Cambridge, Camb, CB2 0QH, UK, E-mail : jl@mrc-lmb.cam.ac.uk

The spliceosome responsible for processing pre-mRNA to mRNA is assembled from RNA-protein particles called snRNPs. The U1, U2, U4 and U5 snRNPs contain in their core a common set of seven Sm proteins (B/B', D1, D2, D3, E, F and G), which assemble on the particle-specific snRNA (small nuclear RNA) by collectively recognizing a conserved U-rich heptad sequence called the Sm site. Addition of particle-specific proteins to the core assembly completes the mature snRNP. The Sm proteins share a conserved sequence motif in two segments, called Sm1 and Sm2, joined by a linker of variable length. We have reconstituted and crystallized the human U4 snRNP core domain comprising the U4 snRNA Sm site with flanking stem-loops and the seven Sm proteins. The crystals belong to space group $P3_1$ ($a = 248.0$ Å, $c = 251.9$ Å) with 12 copies of the core domain in the asymmetric unit. They diffract X-rays to 3.6 Å resolution. The structure was solved by a series of MAD experiments using SeMet-substitution of different groups of Sm proteins, followed by molecular replacement to the native crystal. Our results show that the seven Sm proteins form a closed ring through H-bonding across the Sm1, Sm2 motifs all the way round, and the snRNA is threaded through the pore. The ring has a flat side where the N-terminal helices lie, and a conical side where the linkers protrude. At the rim where the 5' RNA enters the pore from the flat side, the seven Sm proteins bind the Sm site heptad (AUUUUUG) in a one-to-one but asymmetric manner. In addition, the proteins make non-sequence-specific interactions with the 5' and 3' stems. Our findings are applicable to other snRNP cores, because of the conservation of the Sm site and constancy of the Sm proteins.

Keywords: RNA splicing, crystal engineering, noncrystallographic symmetry

MS.08.5

Acta Cryst. (2008). A64, C26-27

Structural basis of lipid biosynthesis regulation in Gram-positive bacteria

Pedro M Alzari¹, Daniela Albanesi¹, Francis Schaeffer¹, Georgina Reh², Alejandro Buschiazzi³, Diego De Mendoza²

¹Institut Pasteur, Structural Biology and Chemistry, 25 rue du Docteur Roux, Paris Cedex 15, Ile de France, 75724, France, ²IBR-CONICET, Facultad de Ciencias Bioquimicas y Farmaceuticas, Universidad Nacional de Rosario, Rosario, Argentina, ³Institut Pasteur de Montevideo, Mataojo 2020, Montevideo, Uruguay, E-mail : alzari@pasteur.fr

FapR is a global transcriptional repressor that controls the expression of several genes involved in the biosynthesis of fatty acids and phospholipids in many Gram-positive bacteria. The FapR monomer has a helix-turn-helix motif linked to an effector-binding domain that displays a 'hot-dog' fold, similar to that of several thioesterases known to process acyl-CoA substrates but different from other known bacterial transcriptional regulators (Schujman et al, 2006). Binding of the specific inducer, malonyl-CoA, promotes conformational changes in the protein that causes the FapR-DNA complex to dissociate or prevents its formation. Furthermore, site-directed mutations which disrupt the FapR-malonyl-CoA interaction result in a lethal phenotype in *Bacillus subtilis*, suggesting that this homeostatic signaling pathway could be a target for novel chemotherapeutic

agents against Gram-positive pathogens. Here we report new binding and structural studies of FapR from *Staphylococcus aureus* in complex with both its DNA operator and effector molecules, providing important hints to understand the mode of action of this conserved bacterial repressor.

Keywords: protein-DNA interactions, regulation of fatty acid biosynthesis, X-ray crystallography

MS.09.1

Acta Cryst. (2008). A64, C27

Seeing the first stages of protein crystal nucleation through to a full powder pattern

Robert Von Dreele

Argonne National Laboratory, Advanced Photon Source, 9700 S. Cass Ave., Argonne, IL, 60439, USA, E-mail: vondreele@anl.gov

The nucleation and growth of protein crystals is the crucial process in any protein structure analysis. Powder diffraction provides a possible means for study of this process because crystallinity can readily be established by diffraction, and a time series of diffraction patterns can be used to explore the crystal growth. Analysis of this data shows not only the time dependence of the amount of crystallized material but also the dimensions of both the crystallites and the crystal unit cell. This talk will show an example of the crystallization of hen egg white lysozyme examined in real time by powder diffraction.

Keywords: powder crystallography, protein crystallization, kinetics of growth

MS.09.2

Acta Cryst. (2008). A64, C27

Serial crystallography: Use of a micro-jet for diffraction of protein nano-crystals or molecules

David A Shapiro^{1,3}, Dan Deponete², Bruce Doak², Petra Fromme², Gary Hembree², Mark Hunter², Stefano Marchesini^{1,3}, Kevin Schmidt², John Spence²

¹Lawrence Berkeley National Laboratory, Advanced Light Source, 1 Cyclotron Road, MS2-400, Berkeley, CA, 94720, USA, ²Arizona State University, PSF470, Tempe AZ, 85287-1504, USA, ³Center for Biophotonics Science and Technology, University of California at Davis, 2700 Stockton Blvd, Suite 1400, Sacramento, CA 95817, USA, E-mail: dashapiro@lbl.gov

We propose a method of acquiring protein powder diffraction data from the smallest available protein crystals that can be acquired through batch precipitation. A continuous micro-jet is used to deliver a solution of hydrated protein nano-crystals to an x-ray beam. This method requires neither the crushing of larger polycrystalline samples nor any techniques to avoid radiation damage such as cryo-cooling. Radiation damage is completely avoided because of the very small dose each crystallite receives as it is only briefly exposed to x-rays. We have commissioned an apparatus to record protein powder diffraction in this manner and in this talk present the first such patterns from photosystem-1 crystals with sizes less than 500 nm. These preliminary patterns show the lowest order reflections, which agree quantitatively with theoretical calculations of the powder profile. The results also serve to test our flow-focusing aerogel injector system, with future application to femtosecond diffraction in Free Electron X-ray Laser schemes, and for Serial Crystallography using a single-file beam of aligned hydrated molecules.

Keywords: serial crystallography, nanocrystals, radiation damage

MS.09.3

Acta Cryst. (2008). A64, C27

Femtosecond laser etching of protein crystal to process and to isolate the single crystal

Yoichiro Hosokawa^{1,2}, Masafumi Kashii^{2,3}, Hiroshi Y. Yoshikawa², Hiroaki Adachi^{2,3}, Yusuke Mori^{2,3}, Hiroshi Masuhara^{1,2}

¹Nara Institute of Science and Technology, Graduate School of Materials Science, 8916-5 Takayama, Ikoma, Nara, 630-0192, Japan, ²Osaka University, 2-1, Suita, Osaka, 565-0871, Japan, ³SOSHO Project (Crystal Design Project), 2-1, Suita, Osaka, 565-0871, Japan, E-mail: hosokawa@hskw.jp

Preparation technique of single protein crystal with suitable shape is needed for precise X-ray diffraction (XRD) measurement giving three-dimensional structure of protein. We have applied an infrared femtosecond laser to isolate and process protein crystal with irregular shape. When the intense femtosecond laser is focused on the protein crystal, electronically excited state of protein molecule is generated densely and the energy is distributed to vibrationally excited states of the molecules in time scale shorter than ns. On the other hand, the time scale for the morphological change of the crystal is longer than ns. Because of the time gap between the photoexcitation energy distribution and the morphological change, a stress is confined at the laser focal point in the time scale shorter than ns. As the result, the etching of the protein crystal by the femtosecond laser is initiated mechanically by the stress increase, though the etching by the other lasers is generally attributed to explosive sublimation explained by conventional heat generation process. The excellence of the femtosecond laser etching is that the cutting is enhanced by a cleaving behavior resulting to stress propagation from the laser focal point. Conclusively the protein crystal has hardly thermal damage by the femtosecond laser irradiation, which was checked by XRD measurement of the small cut component. Furthermore, debris of protein crystal generated by the femtosecond laser etching can be utilized as seed crystal. We confirmed that crystallinity of daughter crystal grew from the seed crystal was fine or comparable in comparison with the mother crystal. These techniques are applicable to select a single crystal from multiple growth forms, even when the size is micrometer-order.

Keywords: protein crystals, pulsed laser ablation, femtosecond laser

MS.09.4

Acta Cryst. (2008). A64, C27-28

Features of the secondary structure of protein molecules from powder diffraction data

Sebastian Basso¹, Celine Besnard¹, Jon P Wright², Irene Margiolaki², Fabrice Camus¹, Gavin C Fox², Andy Fitch², Phil Pattison^{1,3}, Marc Schiltz¹

¹Ecole Polytechnique Federale de Lausanne, Laboratoire de Cristallographie, EPFL - Cubotron IPMC/BSP, lausanne, Vaud, 1015, Switzerland, ²European Synchrotron Radiation Facility (ESRF), ³Swiss-Norwegian Beamlines (SNBL) at ESRF, E-mail: sebastian.basso@epfl.ch

The determination of crystal structures greatly depends upon the availability of good quality single crystals, the growth of which is currently one of the major bottlenecks in macromolecular