

**FA5-MS44-P13**

**Fragment-based Electron Density Interpretation at Low Resolution.** Shao-Yang Ku, Thomas R. Schneider  
*EMBL Hamburg, Germany.*  
E-mail: [s.ku@embl-hamburg.de](mailto:s.ku@embl-hamburg.de)

The electron density of large macromolecular complexes determined by X-ray crystallography and electron microscopy is often only available to low resolution and is difficult to interpret by conventional methods. Fortunately, a complex often contains known structural components, which can be further divided into rigid fragments [1,2]. To position a fragment in an experimentally phased density map at low resolution requires density fitting in real space. Unlike its reciprocal space counterpart, real space molecular replacement has the advantages of ignoring the “missing part” of the structure and allowing the calculation of correlation between the calculated electron density of a search fragment and the experimental electron density within the mask. Here we use the fast spherical averaged density matching as implemented in Molrep [3] to automatically replace fragments into electron density maps between 4 and 10Å resolution, followed by rigid body refinement to maximize the correlation coefficient between the calculated and experimental maps. The method is implemented by using the Clipper libraries [4]. We use the Rab5 GDP/GTP exchange factor (Rabex-5) in complex with human ubiquitin [5] as a test case. The position of the placed ubiquitin fragment is validated by comparing the rmsd between the placed fragment and the refined Rabex-5 structure. We investigate how resolution, phase error, model error and B factors modeling would influence the quality of density fitting.

[1] Schneider T.R. *Acta Cryst. D.* 2004, 60, 2269. [2] Mosca R., Schneider T.R. *Nucleic Acids Res.* 2008, 36, W42. [3] Vagin A.A., Isupov M.N. *Acta Cryst. D.* 2001, 57, 1451. [4] Cowtan K. (2003) IUCr Computing Commission Newsletter, 2, The Clipper C++ libraries for X-ray crystallography, 4-9 [5] Penengo L., Mapelli M., Murachelli A.G., Confaloneri S., Magri L., Musacchio A., Di Fiore P.P., Polo S., Schneider T.R. *Cell* 2006, 124, 1183.

**Keywords:** Low resolution crystallography, electron density fitting, algorithms

**FA5-MS44-P14**

**Validation of B-factor Distributions in Protein Crystal Structures.** Jacopo Negrone<sup>a</sup>, Garib Murshudov<sup>b</sup>, Thomas R. Schneider<sup>a</sup>. <sup>a</sup>*EMBL-Hamburg, Germany.* <sup>b</sup>*YSBL, Chemistry Department, University of York, Heslington, York, England.*  
E-mail: [jacopo.negrone@embl-hamburg.de](mailto:jacopo.negrone@embl-hamburg.de)

Many tools for the analysis of protein models from X-ray crystallography are available nowadays. They check the distribution of geometrical and stereo-chemical properties [1], the agreement of the model with the data [2], or both [3]. These analyses can be either at local or global level. Despite that, a systematic procedure for the analysis and validation of B-factor distributions is still missing. This is surprising since temperature factors play an important role in model interpretation. Moreover, anomalies in the distribution of B-factors can be symptoms of errors introduced during model building and/or refinement. A tool for the detection of these cases would be useful for the interpretation of a protein model

already deposited into the Protein Data Bank (PDB) or at the end of the refinement stage.

Here we propose a new approach for the identification of suspicious B-factor distributions in protein models. The main assumption underlying the method is derived from Bayesian statistics and states that isotropic B-Factors in a protein crystal structure follow an Inverse-Gamma Distribution (IGD). A Maximum Likelihood Estimation (MLE) approach is used to estimate the parameters of the IGD that best fit the distribution of B-factors of a protein structure. A Kolmogorov-Smirnov test (K-S test) is then used to evaluate the goodness of fit and compute a p-value.

We developed and tested the new approach on a set of 14229 protein crystal structures selected from the PDB with a resolution of 2Å or higher. We found that for 82% of the PDB structures the p-value was equal or higher than 0.01, indicating a reasonable agreement between the observed distribution and the expected IGD. For some of the structures with a p-value lower than 0.01, their B-factors still satisfied the IGD assumption if their chains were individually analysed. Thus we analysed only single chains from the original set of PDB structures and we found that around 90% of the chains had a p-value equal or higher than 0.01. Furthermore, a re-refinement protocol performed with the experimental version 5.6 of REFMAC [4] was able to rescue some of the outlier structures found with the single chain analysis.

Our work shows that the IGD distribution is a reasonable assumption for the validation of B-factor distributions and the new approach can be used for the detection of suspicious B-factor distributions in protein models.

[1] Davis IW, Leaver-Fay A, Chen VB, Block JN, Kapral GJ, Wang X, Murray LW, Arendall WB 3rd, Snoeyink J, Richardson JS, Richardson DC., *Nucleic Acids Res.* 2007 Jul; 35(Web Server issue): W375-83. [2] Vaguine AA, Richelle J, Wodak SJ., *Acta Crystallogr D Biol Crystallogr.* 1999 Jan; 55(Pt 1):191-205. [3] Urzhumtseva L, Afonine PV, Adams PD, Urzhumtsev A., *Acta Crystallogr D Biol Crystallogr.* 2009 Mar; 65(Pt 3): 297-300. [4] Murshudov GN, Vagin AA, Dodson EJ., *Acta Crystallogr D Biol Crystallogr.* 1997 May 1;53(Pt 3):240-55.

**Keywords:** structure validation, crystal structure analysis, crystal structure properties

**FA5-MS44-P15**

**A stepwise approach for the X-ray diffraction data in Rietveld refinement.** O.A. Smirnova. *Institute for Chemical Research, Kyoto University, Uji, Kyoto-fu 611-0011, Japan.*

Consideration of the diffraction data in a way they are collected, i.e., with a step applied by a diffractometer, seems a reasonable way to enhance the structure refinement. With this regard, a number of R-factors to evaluate Rietveld fit are suggested. They account for number of points, automatically referring to an equipment resolution. The new R-factors reflect both goodness of background and peaks fitting while conventional Rietveld R-factors neglect background contribution, sometimes making the R-factors artificially high. The true expressions to the R-factors are:

$$R_1 = \frac{\sum(|I_{\text{obs}} - I_{\text{calc}}| / I_{\text{obs}})}{N} \quad R_{1w} = \frac{\sum w_i (|I_{\text{obs}} - I_{\text{calc}}| / I_{\text{obs}})}{N}$$

$$R_2 = \frac{(\sum (|I_{\text{obs}} - I_{\text{calc}}|) / N) / (\sum I_{\text{bragg}} / k_h)}{(\sum w_i |I_{\text{obs}} - I_{\text{calc}}|) / N} \quad R_{2w} = \frac{\sum (w_i |I_{\text{obs}} - I_{\text{calc}}|)}{(\sum w_i I_{\text{bragg}} / k_h)}$$

$$R_3 = \frac{(\sum (|I_{\text{obs}} - I_{\text{calc}}|) / N) / I_{\text{bragg}}}{I_{\text{bragg}}} \quad R_{3w} = \frac{\sum (w_i |I_{\text{obs}} - I_{\text{calc}}|) / N}{w_k I_k}$$

where  $n$  – number of points;  
 $w_i$  – weight,  $w = 1/\sigma$